

Table 1. 400 MHz ^1H NMR methyl group chemical shifts of the C-24 epimeric $\text{C}_{28}\Delta^{5,22}$ -sterols isolated from *Brassica juncea*

	C-18	C-19	C-21	C-26	C-27	C-28
22-Dehydrocampesterol (1)	0.692 (s)	1.009 (s)	1.001 (d, $J = 6.35$)	0.834 (d, $J = 7.33$)	0.817 (d, $J = 6.83$)	0.909 (d, $J = 6.84$)
Brassicasterol (2)	0.692 (s)	1.009 (s)	1.011 (d, $J = 6.84$)	0.833 (d, $J = 6.35$)	0.817 (d, $J = 6.35$)	0.909 (d, $J = 6.84$)

All chemical shifts given in δ -values from TMS; coupling constants in Hz.

were collected each time. GC was carried out on an OV-1 WCOT Si capillary column (0.25 mm i.d. \times 50 m, 280°) and RR_s are given relative to cholesteryl acetate (1.000). ^1H NMR spectra were recorded at 400 MHz with a JEOL JNM FX-400 spectrometer, using CDCl_3 as solvent and TMS as an int. standard.

The $\text{C}_{28}\Delta^{5,22}$ -steryl acetate mixture was obtained from the unsaponifiable matters of *B. juncea* seeds. Prep TLC, gave the sterols which were acetylated and further purified by AgNO_3 -Si gel TLC as described previously [1].

22-Dehydrocampesterol (1). MS m/z (rel. int.): 398 $[\text{M}]^+$ (100), 383 (13), 380 (18), 365 (18), 355 (9), 337 (22), 300 (49), 271 (56), 255 (96), 213 (37). 1-acetate: 380 $[\text{M} - \text{HOAc}]^+$ (100), 365 (9), 337 (10), 282 (9), 255 (62), 228 (11), 213 (16).

Brassicasterol (2). MS m/z (rel. int.): 398 $[\text{M}]^+$ (100), 383 (16), 380 (19), 365 (18), 355 (11), 337 (28), 300 (56), 271 (65), 255 (94), 213 (34). 2-acetate: 380 $[\text{M} - \text{HOAc}]^+$ (100), 365 (9), 337 (10), 282 (9), 255 (61), 228 (11), 213 (15).

Acknowledgements—We thank K. Matsumoto and H. Yazima for

technical assistance, Dr. Y. Fujimoto for running the 400 MHz ^1H NMR spectra.

REFERENCES

1. Matsumoto, T., Shimizu, N., Itoh, T., Iida, T. and Nishioka, A. *J. Am. Oil Chem. Soc.* (in press).
2. Ashikaga, C. (1957) *Nippon Nogei Kagaku Kaishi* **31**, 115.
3. Bolker, H. I. (1967) *Nature (London)* **213**, 905.
4. Rubinstein, I. and Goad, L. J. (1974) *Phytochemistry* **13**, 485.
5. Kircher, H. W. and Rosenstein, F. U. (1973) *Lipids* **8**, 453.
6. Teshima, S., Patterson, G. W. and Dutky, S. R. (1980) *Lipids* **15**, 1004.
7. Morris, R. J. and Culkin, F. (1977) *Oceanogr. Mar. Biol. Annu. Rev.* **15**, 73.
8. Khalil, M. W., Idler, D. R. and Patterson, G. W. (1980) *Lipids* **15**, 69.
9. Chiu, P.-L. and Patterson, G. W. (1981) *Lipids* **16**, 203.
10. Rubinstein, I., Goad, L. J., Clague, A. D. H. and Mulheirn, L. J. (1976) *Phytochemistry* **15**, 195.

A XYLOSYLGLUCOSIDE OF XANTHOXYLIN FROM *SAPIUM SEBIFERUM* ROOT BARK

ISAO KOUNO, TOSHIHIRO SAISHOJI, MIDORI SUGIYAMA and NOBUSUKE KAWANO*

Faculty of Pharmaceutical Sciences, Nagasaki University, 1-14 Bunkyo-machi, Nagasaki 852, Japan

(Received 11 August 1982)

Key Word Index—*Sapium sebiferum*; Euphorbiaceae; root bark; moretenone; moretenol; xanthoxylol; sitosterol β -D-glucoside; 2-acetyl-3,5-dimethoxyphenyl-O- β -D-xylopyranosyl-(1-6)- β -D-glucopyranoside; ^1H NMR; ^{13}C NMR.

Abstract—Besides four known compounds, a new xylosylglucoside of xanthoxylol was isolated from the root bark of Chinese tallow tree, *Sapium sebiferum* and identified as 2-acetyl-3,5-dimethoxyphenyl-O- β -D-xylopyranosyl-(1-6)- β -D-glucopyranoside.

INTRODUCTION

Chinese tallow tree, *Sapium sebiferum* Roxb. is common in Nagasaki city as a roadside tree. Its root bark has been

used [1] in China as a purgative and diuretic. Recently, it was reported [1] to be effective against *Schistosoma japonicum*. However, only xanthoxylol (2-hydroxy-4,6-dimethoxyacetophenone) has been isolated from the root bark [2] although some other constituents, such as

* Author to whom correspondence should be addressed.

friedelin, sitosterol and ellagic acid from leaves and moretenone, moretenol and sterols from stem have been reported [3]. The present paper describes the isolation and structure elucidation of a xylosylglucoside obtained from the root bark.

RESULTS AND DISCUSSION

The methanol extracts of the bark were fractionated with chloroform, ethyl acetate, and *n*-butanol successively. Moretenone, moretenol [4], xanthoxylin (1), and sitosterol- β -D-glucoside were obtained from the chloroform-soluble part. The *n*-butanol-soluble part yielded a xylosylglucoside (2), mp 188–190°, which afforded 1, xylose and glucose on acid hydrolysis. Compound 2 gave a hexa-acetate 3 and a hexamethyl ether 4. The mass spectra of 3 and 4 suggested the presence of a terminal xylose because their base peaks, observed at *m/z* 175 and 259, are assignable to xylose fragments. The ^{13}C NMR spectrum of 2 suggested a xylo-(1-6)-glucoside structure because two methylene carbon signals at δ 66.9 and 70.1 are assignable [5] to C-5 of xylose and glycosylated C-6 of glucose, respectively. Final confirmation was obtained by methanolysis of 4 followed by GC detection of 1, methyl-2,3,4-tri-*O*-methyl-D-xylopyranoside, and methyl-2,3,4-tri-*O*-methyl-D-glucopyranoside. In the ^1H NMR spectrum of 4, two anomeric proton signals observed at δ 4.20 and 4.80 are doublets ($J = 7.2$ Hz), suggesting β -configurations in both the sugar linkages. Therefore, the structure of the glycoside was deduced as 2-acetyl-3,5-dimethoxyphenyl-*O*- β -xylopyranosyl-(1-6)- β -D-glucopyranoside.

EXPERIMENTAL

General. Mps are uncorr. Shimadzu GC-3BF gas chromatograph equipped with a column (2.1 m) of 10% SE-30 was used for GC. Kiesel gel 60 (Merck) and Kiesel gel 60 GF₂₅₄ were used for CC and TLC, respectively.

Isolation. The fresh root bark (2 kg) of Chinese tallow tree collected in Oct. at Nagasaki was ground with MeOH and extracted $\times 3$ at room temp. The resulting MeOH extracts (64 g) were fractionated with CHCl_3 , EtOAc and *n*-BuOH, successively. The CHCl_3 -soluble part (9 g) was separated by CC (Si gel) eluting with a mixture of *n*-hexane and EtOAc to give moretenone (152 mg), xanthoxylin (327 mg), moretenol (5.3 mg) and sitosterol- β -D-glucoside (53 mg). The *n*-BuOH-soluble part (16 g) was dissolved in H_2O and separated by CC (Amberlite XAD-2) eluting with H_2O , H_2O -MeOH and MeOH, successively. The latter two fractions gave a xanthoxylin glycoside (290 mg) after a further CC (Si gel, CHCl_3 -MeOH, 4:1).

Moretenone. Colourless needles from MeOH, mp 196–198°, MS *m/z* (rel. int.): 424 [M]⁺ (90), 409 (98), 381 (16), 205 (100); ^1H NMR (CDCl_3): δ 0.70 (3H, s), 0.95, 1.03 (6H, s each), 1.08, 1.68 (3H, s each), 4.68 (2H, br s). (Found: C, 84.73; H, 11.43. $\text{C}_{30}\text{H}_{48}\text{O}$ requires: C, 84.84; H, 11.39%.)

Moretenol. Colourless scales from Me_2CO , mp 215–217°, MS *m/z* (rel. int.): 426 [M]⁺ (80), 411 (28), 383 (15), 368 (48), 207 (100); ^1H NMR (CDCl_3): δ 0.69, 0.72, 0.83, 0.94 (3H, s each), 0.97 (6H, s), 1.67 (3H, s), 3.51 (1H, s), 3.84 (1H, d, $J = 2.7$ Hz), 4.68 (2H, br s). PCC oxidation gave moretenone (TLC, MS and ^1H NMR).

Xanthoxylin (1). Colourless needles from MeOH, mp 77–79°, ^1H NMR (CDCl_3): δ 2.59, 3.80, 3.83 (3H, s each), 5.88, 6.03 (1H, d, $J = 2$ Hz each), 13.92 (1H, s).

Xanthoxylin glycoside (2). Colourless needles from MeOH, mp 188–190°, $[\alpha]_{\text{D}}^{25} -41.7^\circ$ (pyridine; 0.5%); UV $\lambda_{\text{max}}^{\text{EtOH}}$ (e) nm: 225 (14 520) and 273 (7570); IR $\nu_{\text{max}}^{\text{nujol}}$ cm^{-1} : 1645, 1600, 815, 800; ^1H NMR (d_5 -pyridine): δ 2.74, 3.64, 3.79 (3H, s each), 5.52 (1H, d, $J = 7.2$ Hz), 6.33, 7.02 (1H, d, $J = 1.8$ Hz each); ^{13}C NMR (d_5 -pyridine): δ 32.9 q, 55.7 q, 55.9 q, 66.9 t, 70.1 t, 71.0 d, 71.3 d, 74.7 d, 74.8 d, 77.5 d, 78.0 d, 78.3 d, 93.8 d, 95.7 d, 103.5 d, 105.8 d, 116.1 s, 157.1 s, 158.4 s, 162.8 s, 201.0 s. (Found: C, 50.88; H, 6.20. $\text{C}_{21}\text{H}_{30}\text{O}_{13}$ requires: C, 51.42; H, 6.17%.) Hexa-acetate of 2: Compound 2 (50 mg) was acetylated with Ac_2O (2 ml) and pyridine (4 ml) and purified by CC (Si gel, *n*-hexane-EtOAc, 4:1) to give colourless plates (45 mg) from 50% MeOH, mp 128–130°, MS *m/z* (rel. int.): 742 [M]⁺ (6.5), 684 (2.9), 548 (48), 368 (7.2), 331 (4.3), 317 (34), 280 (17), 259 (100, VI), 215 (14); ^1H NMR (CDCl_3): δ 1.66, 1.96, 2.01, 2.06, 2.06, 2.08, 2.40, 3.79, 3.82 (3H, s each), 3.2–3.6 (3H, m), 4.0–4.3 (3H, m), 4.52 (1H, d, $J = 6.3$ Hz), 4.8–5.3 (6H, m), 6.23, 6.32 (1H, d, $J = 1.8$ Hz each). (Found: C, 53.37; H, 5.88. $\text{C}_{33}\text{H}_{42}\text{O}_{19}$ requires: C, 53.37; H, 5.70%.)

Acid hydrolysis of 2. A mixture of 2 (30 mg) and 3% H_2SO_4 (4 ml) was refluxed for 1.5 hr and extracted with CHCl_3 to give a solid (6.3 mg), which was identified with 1 (MS and ^1H NMR). The aq. layer was neutralized with $\text{Ba}(\text{OH})_2$ and the filtrate separated from BaSO_4 was analysed by avicel TLC for detection of xylose (R_f : 0.47 and 0.34) and glucose (R_f : 0.39 and 0.23); solvents used: pyridine-EtOAc-HOAc- H_2O (36:36:7:21) and *n*-BuOH-HOAc- H_2O (3:1:1), respectively.

Methanolysis of the hexamethyl ether of 2. Compound 2 (30 mg) in DMF (1 ml) was mixed with MeI (0.5 ml) and Ag_2O (1.3 g). The mixture, in a sealed tube, was kept at 37° for 6 hr and then left standing overnight at room temp. The reaction mixture was extracted with CHCl_3 and the CHCl_3 -soluble part was once more methylated by the same way as above except for the use of BaO (800 mg) instead of Ag_2O [6]. The product was purified by CC (Si gel, *n*-hexane-EtOAc, 7:3) to give an oil (7.5 mg). MS *m/z* (rel. int.): 574 [M]⁺ (3), 531 (3), 391 (20), 378 (34), 370 (48), 313 (26), 279 (26), 256 (23), 196 (64), 175 (100, V); ^1H NMR (CDCl_3): δ 2.47, 3.45, 3.47, 3.56, 3.56, 3.59, 3.63, 3.79, 3.79 (3H, s each), 4.20, 4.80 (1H, d, $J = 7.2$ Hz each), 6.15, 6.32 (1H, d, $J = 2.7$ Hz each). The methyl ether (3.5 mg) was mixed with 0.3 N HCl-MeOH (5 ml) and refluxed for 2 hr. The cooled soln was neutralized with Ag_2CO_3 and the filtrate was evaporated to dryness. The reaction product was dissolved in Me_2CO and subjected to GC for detection of the following compounds: methyl-2,3,4-tri-*O*-methyl- β -D-xylopyranoside, R_t 1.9 min; α -compound, R_t 2.4 min; methyl-2,3,4-tri-*O*-methyl- β -D-glucopyranoside, R_t 4.6 min; α -compound, R_t 5.2 min; xanthoxylin (1), R_t 10.8 min. Each compound was detected by co-chromatography with an authentic sample.

REFERENCES

1. Chu, Y. L., Hsu, C. L. and Liu, P. S. (1958) *Yao Hsueh Hsueh Pao* 6, 51.
2. Liu, C. T. and Loh, J. Y. (1957) *Yao Hsueh Hsueh Pao* 23, 259.
3. Hui, W. H., Fung, T. L. and Ng, K. K. (1969) *Phytochemistry* 8, 331.
4. Galbraith, M. N., Miller, C. J., Rawson, J. W. L., Ritchie, E., Shannon, J. S. and Taylor, W. C. (1965) *Aust. J. Chem.* 18, 226.
5. Konoshima, T., Fukushima, H., Inui, H., Sato, K. and Sawada, T. (1981) *Phytochemistry* 20, 139.
6. Lloyd, K. O., Kabat, E. A., Layug, E. J. and Gruezo, F. (1966) *Biochemistry* 5, 1489.